

Tracking settings with Rainbow Beads

Purpose: to ensure consistent signals over the course of a project, and verify proper function of the cytometers.

Rationale: The components of a cytometer can display variability from day to day. Fluctuations in laser power, changes in temperature and humidity affecting the PMT's, and changes in alignment on the instrument are just some examples of variables that can affect the output. In order to minimize these variations, we should apply a consistent fluorescence standard with each experiment to make sure that the signal output of the instrument for a given level of fluorescence is consistent. To do this, we use a stable source of fluorescence - [Rainbow Calibration Particles \(8 peaks\)](#)

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[Ref: 559123](#) Enzifarma

A mixture of 3.0 - 3.4 μm Rainbow Particles that are dyed to eight different fluorescent intensities. Every Rainbow Particle contains a mixture of fluorophores that are excited at any wavelengths from 365 - 650 nm.

Procedure: After establishing settings for a given panel of reagents such that all signals are on scale, compensation between channels is minimized, and separation of populations is either optimal (for dim labels) or sufficient (for bright ones), the operator collects a data file of rainbow beads that gives positive signals for the established settings. For the next experiment, the first step would be to run the rainbow beads, and make sure that the uncompensated MFIs are $<\pm 15\%$ different from the initial run.

Step by step

On the initial day of your experiment

Establish your panel settings and run your experiment (set voltages; calculate compensation; collect your data files);



“Duplicate without Data” to create a copy of the experiment you just ran. Call it [Setup Experiment](#).

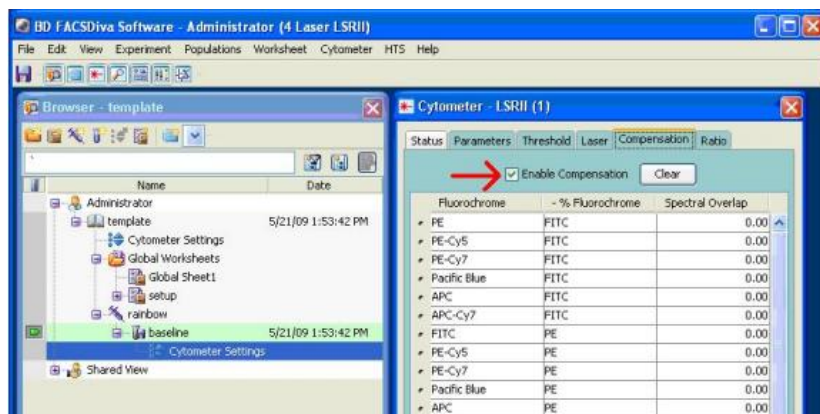


Delete the specimen and tubes from the original experiment and create a new specimen. Then create the first tube, call it “[baseline](#)” and collect a data file of 8-peak rainbow beads (It may be necessary to change the FSC or SSC settings to be able to visualize the beads).



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Select “Cytometer settings” to view the Compensation tab in the inspector. Clear all the compensation values or de-select the “Enable compensation” bottom.



On the global worksheet, draw a FSC/SSC dot plot, and a histogram for each color.



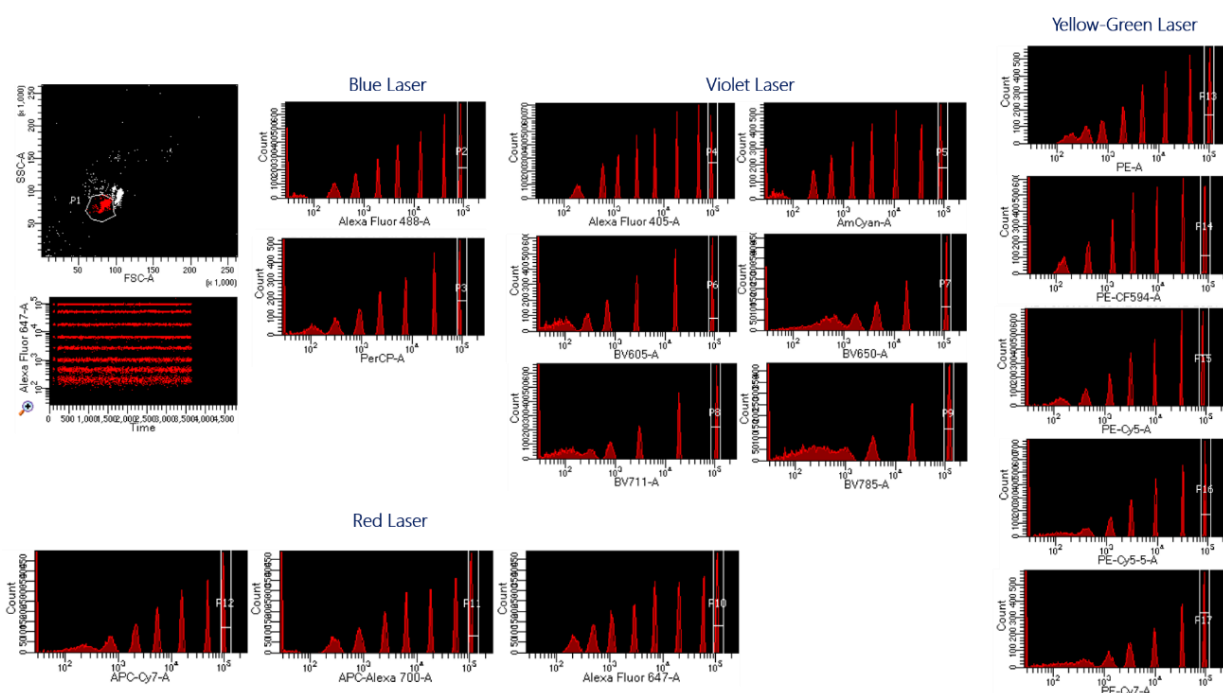
On each histogram, draw interval gates on the 8th or 7th peak and evaluate the MFI for each one of the colors

(Ask for the Statistics Table, right clicking on a plot; Edit Statics, right clicking on the statistics table and ask for the Mean value for all the colors).



Export the experiment and save the template.

Rainbow Calibration beads



STANDARD OPERATING PROCEDURES

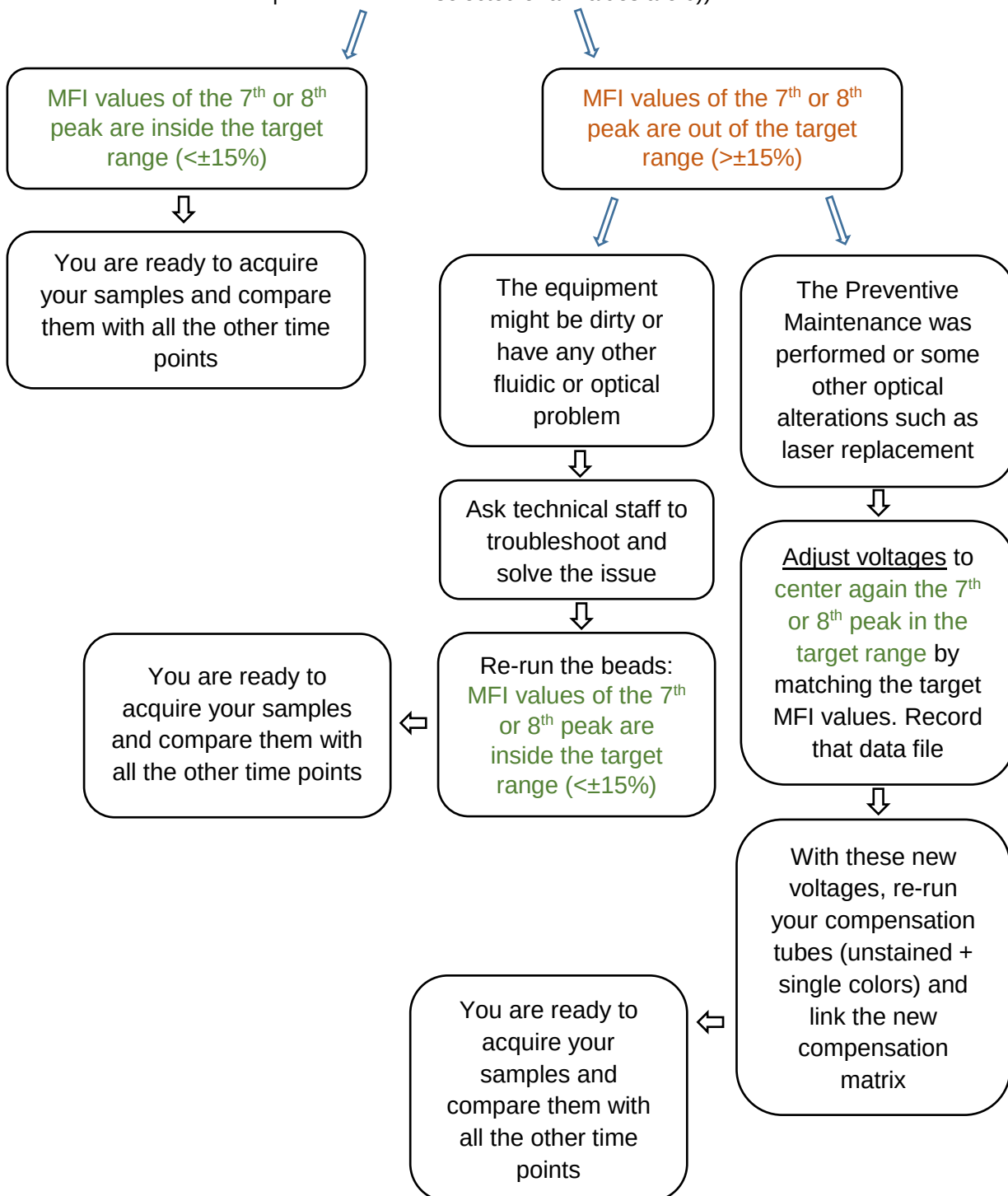
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On subsequent experiments

Open the *Setup Experiment*



“Next Tube”, label with the date, run and record the 8-peak Rainbow beads.
(Every time, check the instrument settings to make sure compensation is disabled (“Enable compensation” is de-selected or all values are 0)).



**SOP.UCF.015 – TRACKING SETTINGS
WITH RAINBOW BEADS**

Reference peak target MFI				
Date:			RB LOT #	
Channel	Lower MFI (-15%)	Target MFI	Upper MFI (+15%)	Rainbow bead peak #

Reference peak target MFI				
Date:			RB LOT #	
Channel	Lower MFI (-15%)	TARGET MFI	Upper MFI (+15%)	Rainbow bead peak #